

Nab-Paclitaxel-Loaded Poly (Lactic-Co-Glycolic Acid) Nanoparticles as Microtubule B-Tubulin Stabilizer in the Management of Pancreatic Cancer

^{1b} Bryan Gervais de LIYIS^a, ^{1b} Viona MARESKA^a, ^{1b} Celine Aurelia AHMAD^a,
^{1b} Alexandria Cahyaputri WINDIARTO^a, ^{1b} Agung Wiwiek INDRAYANI^b

^aUdayana University Faculty of Medicine, Bali, Indonesia

^bDepartment of Pharmacology and Therapy, Udayana University Faculty of Medicine, Bali, Indonesia

ABSTRACT Pancreatic cancer, specifically driven by the Kirsten rat sarcoma virus gene mutation (KRAS), remains a formidable clinical challenge with limited therapeutic options. The absence of FDA-approved drugs directly targeting KRAS necessitates exploration of novel and more effective treatment strategies. This comprehensive literature review seeks to identify promising therapeutic avenues for pancreatic cancer by evaluating advancements in drug delivery systems. Nab-paclitaxel, an antimetastatic agent, exhibits superior pharmacokinetic and bioavailability profiles compared to conventional paclitaxel. Utilizing poly (lactic-co-glycolic acid) (PLGA) nanoparticles as carriers, we investigate the potential of nab-paclitaxel-loaded PLGA nanoparticles to enhance drug delivery and efficacy. Manufactured through the oil-in-water emulsification solvent evaporation method, nab-paclitaxel-loaded PLGA nanoparticles offer a faster half-life and undergo elimination via biliary excretion and metabolism. Notably, these nanoparticles leverage nanoalbumin interactions with cysteine/osteonection-rich, acidic secreted proteins, resulting in highly selective targeting of pancreatic cancer cells. The findings of this review underscore the potential superiority of nab-paclitaxel-loaded PLGA nanoparticles in terms of pharmacokinetics, pharmacodynamics, and clinical outcomes. Their ability to address the challenges posed by KRAS-driven pancreatic cancer holds promise as a transformative approach in the treatment landscape. In conclusion, this review highlights the evolving landscape of therapeutic options for pancreatic cancer, shedding light on nab-paclitaxel-loaded PLGA nanoparticles as a potent and selective intervention. Further clinical validation and exploration of this innovative strategy are warranted to advance the management of this devastating disease.

Keywords: Nab-paclitaxel; nanoparticles; pancreatic cancer; polylactic acid-polyglycolic acid copolymer

Pancreatic cancer has a poor prognosis, with a survival rate of approximately 9%.¹ It is the 14th most common cancer and the 7th leading cause of cancer deaths, with a total of 458,918 cases and 432,242 deaths globally in 2018.² The study by Saad et al. emphasized an increase in the pancreatic cancer prevalence by 1.03% annually from 1973 to 2014 and pancreatic cancer was predicted to be the second leading cause of death by 2030.³ The pancreatic cancer prevalence increased by 39.53% at 55-64 years of age and more than 95% of cases had metastasized at the time of diagnosis or in the future.^{4,5} This is a manifestation of mutations in the dominant isoform that is mutated in pancreatic cancer, that is, the Kirsten rat sarcoma virus (KRAS) gene sequence.⁶

KRAS functions as a molecular switch that initiates intracellular signaling pathways and transcription factors inducing cell proliferation, migration, transformation, and survival, and mutations lead to uncontrolled cell growth leading to cancer.⁷

The first-line treatment of advanced pancreatic cancer has been single-agent gemcitabine for more than a decade.⁸ Deoxycytidine kinase activates nucleoside diphosphate and triphosphate, leading to deoxyribonucleic acid (DNA) polymerase competitive inhibition, thus, impeding DNA synthesis through intracellular conversion by gemcitabine.⁹ Chemotherapeutic agents have low efficacy and chemoresistance and prolong patient survival by less than one year.¹⁰⁻¹³ Moreover, gemcitabine has certain side effects, in-

TO CITE THIS ARTICLE:

de Liyis BG, Mareska V, Ahmad CA, Windiaro AC, Indrayani AW. Nab-Paclitaxel-Loaded Poly (Lactic-Co-Glycolic Acid) Nanoparticles as Microtubule B-Tubulin Stabilizer in the Management of Pancreatic Cancer. Journal of Oncological Sciences. 2023;9(3):166-75.

Correspondence: Bryan Gervais de LIYIS
Udayana University Faculty of Medicine, Bali, Indonesia
E-mail: bryan.gervais@student.unud.ac.id

Peer review under responsibility of Journal of Oncological Sciences.

Received: 31 Oct 2022

Received in revised form: 04 Apr 2023

Accepted: 18 Jul 2023

Available online: 11 Sep 2023

2452-3364 / Copyright © 2023 by Turkish Society of Medical Oncology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).



cluding heart toxicity, peripheral edema, nausea, vomiting, and anorexia.^{12,14,15} No current Food and Drug Administration (FDA) approved drugs directly target the KRAS mutation proteins.¹⁶ This is owing to the challenges in the development of small molecule inhibitors that have a sufficiently high affinity for KRAS mutations. Moreover, KRAS mutations have an extremely high affinity for guanosine triphosphate (GTP), and their catalytic sites are small and difficult to target.¹⁷ Additionally, the KRAS gene mutation impact can be inhibited by preventing the replication of mutated cells using antimetabolic drugs, such as paclitaxel.¹⁸

The most significant advancement in chemotherapy for breast, endometrial, non-small cell lung, bladder, and cervical cancers in the last two decades is paclitaxel, the first identified microtubule stabilizing agent.¹⁹ However, paclitaxel has a limited clinical application owing to its hydrophobicity and low therapeutic index.²⁰ Nonetheless, this limitation can be avoided through the use of new drug delivery systems such as poly (lactic-co-glycolic) acid (PLGA) paclitaxel nanoparticles with sizes up to 200 nm.^{20,21} PLGA is an effective biodegradable polymer nanoparticle drug delivery system owing to its controlled and sustained release, low levels of toxicity, and tissue biocompatibility.²² PLGA nanoparticles are formulated from biocompatible and biodegradable polymers and are used in research owing to their small size distribution, synthesis with controlled reaction time and temperature, high structural integrity, and high production rate.²³ The systemic delivery of insoluble compounds to tumor sites is facilitated by the encapsulation of various hydrophobic chemotherapeutics by PLGA nanoparticles.^{21,24} An increased intratumoral concentration mediated by 60 kDa albumin-binding glycoprotein (gp60) is associated with increased tumor response to nab-paclitaxel, facilitating vascular transcytosis.²⁵

Thus, this review aimed to determine better prospects regarding the management of pancreatic cancer by acknowledging the potential of paclitaxel and the use of PLGA nanoparticle technology.

PANCREATIC CANCER

The pancreas is a metabolic organ with both exocrine and endocrine roles.²⁶ The exocrine glands comprise

pancreatic acinar and duct cells that induce digestive enzymes and sodium bicarbonate production, respectively.²⁷ Endocrine glands consist of 5 different types of secretory islet cells that produce peptide hormones to control glucose levels.²⁸ Pancreatic cancer arises from genetic defects and causes abnormal cell proliferation and function.²⁹ In pancreatic cancer, there is a loss and gain of chromosomes.³⁰ The chromosome arm 9p21, 17p13, 18q21, 3p, 8p22, and 6q are the most prevalent areas of genome loss in primary pancreatic cancer.³¹⁻³⁶ Genes related to tumor-suppressing, such as cyclin-dependent kinase inhibitor 2A (CDKN2A)/P16/MTS1 (in 9p21), p53 (in 17p13), and mothers against decapentaplegic homolog 4/SMAD family member 4 (SMAD4)/deleted in pancreatic cancer 4 (in 18q) are found in these loci.³⁷⁻⁴⁰ Fluorescent in-situ hybridization demonstrated increased DNA regions at 12p, 12q, 17q, 19q, and 20q.⁴¹ The amplified sites are consistent with the locations of the oncogenes AKT serine/threonine kinase 2 (AKT2) (in 19q), KRAS (in 12p), mouse double minute 2 homolog (in 12q), Erb-b2 avian erythroblastic leukemia viral oncogene homolog 2 (in 17q) and amplified in breast cancer 1 (in 20q).⁴²⁻⁴⁸ KRAS, the most frequently mutated oncogene (90%) in pancreatic cancer, is located on the 12p arm of the chromosome.⁴⁹ A meta-analysis of 34 studies on P53, SMAD4, CDKN2A/P16, and KRAS, containing 3373 samples, found that KRAS (hazard ratio=1.68, 95% confidence interval=1.27-2.22, $p<0.001$) had the highest mutational significance in pancreatic cancer compared to other oncogenes.⁵⁰

KRAS

A key component of cellular networks is the RAS protein that governs growth, proliferation, survival, differentiation, adhesion, cytoskeletal rearrangement, and cell motility through a variety of signaling pathways (Figure 1).⁵¹ Three members of the RAS gene family, which includes Harvey rat sarcoma virus, KRAS, and neuroblastoma rat sarcoma virus, act as proto-oncogenic factors in human tumor activation.⁵² The KRAS mutation is an early genetic event for pancreatic cancer.⁵³

KRAS can only bind and activate effector proteins like rapidly accelerated fibrosarcoma (Raf)-ki-

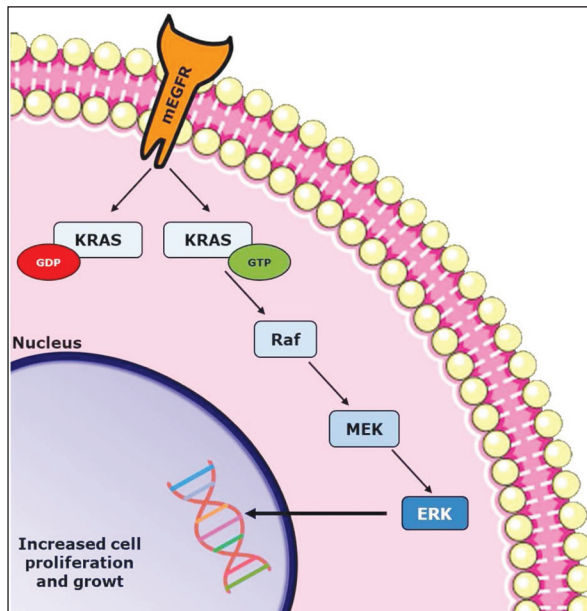


FIGURE 1: Ras/Raf/MEK/ERK pathway.

Ras: Rat sarcoma virus; Raf: Rapidly accelerated fibrosarcoma; MAPK: Mitogen-activated protein kinase; ERK: Extracellular signal-regulated kinase; mEGFR: Mutant epidermal growth factor receptor; KRAS: Kirsten rat sarcoma virus; GDP: Guanosine diphosphate; GTP: Guanine triphosphate.

nase, phosphoinositide 3-kinase, and Ral guanine nucleotide dissociation stimulator when it is GTP-bound.⁵⁴ KRAS is activated when the protein guanine exchange factor supersedes guanosine diphosphate from the nucleotide-binding site owing to higher intracellular GTP concentrations.⁵⁵ KRAS signaling hyperactivity, which can occur through direct KRAS mutation or indirectly through other proteins in the KRAS pathway, leads to Raf activation in the Ras/Raf/MEK/ERK pathway.^{56,57} This pathway plays an important role in tumor cell survival and development.⁵⁸ This subsequently causes uncontrolled cell proliferation. Several attempts have led to the identification of compounds that specifically block important factors for mitosis in cancer therapy.

NAB-PACLITAXEL AND POLY (LACTIC-CO-GLYCOLIC) ACID

Antimitotic chemotherapeutic agents, such as vinblastine and taxol, halt the cell mitotic cycle progression, causing cancer cells to undergo apoptosis.⁵⁹ The microtubules create spindles during prophase to draw the chromosomes toward the poles. During the later

stages, the microtubules undergo depolymerization, dissolving the structure.⁶⁰ Paclitaxel binds and stabilizes microtubules, preventing depolymerization (Figure 2).⁶¹ Therefore, tubulin polymerization is promoted, and mitotic progression is inhibited by paclitaxel.⁶² Despite using paclitaxel in the management of various cancers, including cervical, breast, ovarian, bladder, prostate, liver, and lung, the clinical application of paclitaxel is highly limited owing to its efficacy and multi-drug resistance (MDR) properties.⁶³ Nanoparticles have become the treatment of choice for cancer owing to their good pharmacokinetics, precise targeting efficacy, reduction of side effects, and more significant anti-drug resistance without increasing therapeutic hazard to patients.^{64,65}

Nab-paclitaxel is a type of taxane that interferes with cell division and prevents the growth and spread of cancer cells.⁶⁶ Unlike conventional paclitaxel, formulated using a solvent called Cremophor EL (Sigma Chemical Co., St. Louis, Missouri, United States) that can cause severe allergic reactions and other side effects in some patients, nab-paclitaxel is formulated with albumin-bound nanoparticles delivering the

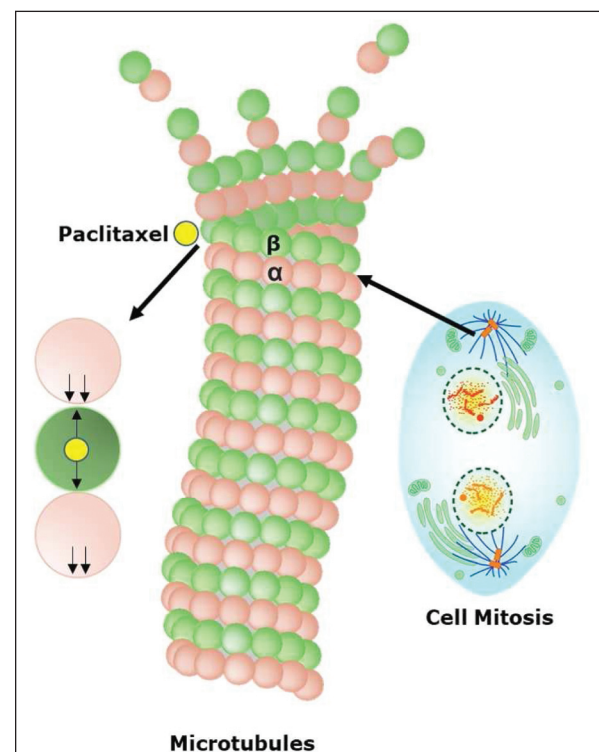


FIGURE 2: Paclitaxel as β -tubulin stabilizer in cell microtubules.

drug more efficiently to the tumor cells.⁶⁶ These nanoparticles protect the drug from degradation and clearance (CL) by the body's immune system, allowing for more drugs to reach the tumor cells.^{67,68} Furthermore, the nanoparticles can bind to a specific protein found on the surface of tumor cells, enhancing the drug's ability to enter the cells and exert its therapeutic effects.⁶⁸ Nab-paclitaxel has a higher response rate and improved progression-free survival (PFS) compared to conventional paclitaxel. For example, in a Phase III clinical trial comparing nab-paclitaxel to conventional paclitaxel in patients with metastatic breast cancer, nab-paclitaxel was associated with a higher overall response rate (ORR) (33% vs. 19%) and longer PFS (23.0 vs. 16.9 weeks).⁶⁹ The use of albumin-bound nanoparticles in nab-paclitaxel mitigates the potential occurrence of allergic reactions and other adverse effects. Notably, nab-paclitaxel is associated with lower incidence rates of all-grade neuropathies, anemia, pain, and diarrhea than paclitaxel, while also significantly reducing the use of antiemetics and antihistamines.^{70,71} The unique formulation of nab-paclitaxel with albumin-bound nanoparticles improves the drug's efficacy while reducing its side effects, thus, making it a superior chemotherapeutic drug compared to conventional paclitaxel.

However, the clinical utilization of paclitaxel is restricted by its low solubility, prompting the exploration of alternative delivery methods. Thus, PLGA nanoparticles, which are modified using tocopheryl polyethylene glycol succinate (TPGS) and vitamin E via solvent evaporation, have been utilized to encapsulate paclitaxel and control its release *in vitro*, to address this limitation.⁷² PLGA, an FDA-approved synthetic polymer, shows great promise as a drug delivery vehicle owing to its biodegradability, biocompatibility, and ease of surface modification for site-specific drug release.⁷³ Nanoparticle encapsulation of anticancer drugs enhances release rates while reducing the risk of toxicity owing to the large surface area-to-volume ratio and targeted delivery.⁷⁴ According to a study, the atomic force microscopy findings indicated no alterations in the physical properties of PLGA nanoparticles after loading paclitaxel in pancreatic cancer cells, which was supported by a similar modulus in paclitaxel-loaded PLGA

nanoparticles of approximately 12 GPa; thus, verifying the feasibility of loading paclitaxel into PLGA.²¹ Previous clinical trial indicates that the use of polyethyleneimine-formulated PLGA nanoparticles results in increased cellular uptake, sustained siRNA, and effective gene deliveries.⁷⁵ In contrast to conventional liposomes with a half-life of less than 10 h, PEGylated PLGA liposomes demonstrate a longer elimination half-life of approximately 50 h, resulting in higher intratumoral paclitaxel accumulation and improved antitumor efficacy in mice with colon-26 solid tumor-bearing.^{76,77} The existing literature suggests that PLGA in combination with nab-paclitaxel could potentially yield considerable therapeutic benefits, despite the limitations of current clinical trials investigating the efficacy of this integration.

MECHANISM OF CONSTRUCTION AND ADMINISTRATION OF NAB-PACLITAXEL-LOADED PLGA

To synthesize PLGA nanoparticles comprising biomacromolecules, the oil-in-water (o/w) emulsification solvent evaporation method has become the method of choice owing to its more preferable requirements. Firstly, PLGA is dissolved in an organic, volatile, hydrophobic solvent such as dichloromethane (DCM). Next, the mixture is emulsified in a large amount of liquid in the aqueous phase using an ultrasound or a high-velocity homogenizer and an emulsifier or surfactant, most commonly polyvinyl alcohol (PVA) or TPGS. Afterward, organic solvent from the mixture is eliminated to create particles either by using low-pressure evaporation or dilution with large volumes of water or some other cooling agents to spread the solvent out. Lastly, the excess PVA or TPGS is removed and freeze-dried by rinsing the solid particles.⁷⁸

The nab-paclitaxel-loaded nanoparticles are prepared using the o/w emulsification solvent evaporation technique (Figure 3). Firstly, 5 mg of paclitaxel is dissolved in a flask containing 4 mL of DCM with a 2.5% (w/v) PLGA. The resultant product was put into 20 mL of 1% (w/v) TPGS while stirring and sonicating. A prepared magnetic stir plate should swirl the remaining emulsion continuously at 25°C for approximately 6 h to evaporate the DCM. Following this, produced nanoparticles will be centrifuged and

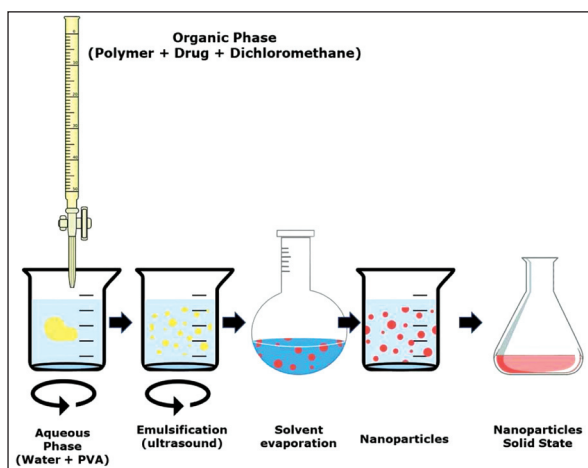


FIGURE 3: Schematic representation of the oil-in-water emulsification of solvent evaporation preparation method.

PVA: Polyvinyl alcohol.

rinsed with distilled water multiple times. The nanoparticles are then lyophilized and stored at 4°C before the next use.^{79,80} Additionally, the morphology of the nanoparticles is determined using scanning electron microscopy, and the size of the nanoparticles on the electron micrograph is assessed using Adobe Photoshop (Adobe Systems, San Jose, California, United States).⁸¹ Afterwards, 5 mg of paclitaxel-loaded nanoparticles are dissolved in 1 mL of DCM. Then, 3 mL of a 50:50 v/v acetonitrile: water mixture is added and extracted. A continuous nitrogen stream is used to evaporate DCM, which will then result in a clear translucent solution. High-performance liquid chromatography (HPLC) equipment is used to identify the composition of the resulting mixture. The HPLC test is performed at a flow rate of 1.0 mL/min with a 50:50 (v/v) combination of acetonitrile and water. A variable wavelength detector is used to detect paclitaxel. The calibration curve for paclitaxel quantification can then be linear across the standard range between 50 to 100,000 ng/mL with a correlation value of $R^2=1.0$.⁸²⁻⁸⁵

The recovery efficiency factor of the extraction procedure of encapsulation efficiency (EE) was determined to assess a certain weight of pure paclitaxel. Approximately 5 mL of phosphate-buffered saline (PBS) was added. The extraction technique as described previously was used and the drug's efficiency was then determined as a result.⁸⁶ The following formulae are used to know the loading efficiency and EE:

Loading efficiency (%)=(amount of drug in nanoparticles/number of nanoparticles loaded with drug) $\times 100$ EE (%)=(amount of drug in nanoparticles/initial amount of drug) $\times 100$. A deep HPLC test in PBS with a pH of 7.4 was used to determine the amount of nab-paclitaxel released from nanoparticles. Approximately 5 mg of paclitaxel-charged nanoparticles are suspended in 10 mL of PBS in a sealed tube and shaken at 37°C and 120 rpm in an orbital shaker. The supernatant is then suspended in 1 mL of DCM to obtain the nab-paclitaxel concentration.⁸⁶⁻⁸⁹ After weighing the syringe to reach the desired dosage of paclitaxel 10 mg/kg, the preparation is delivered with a dose volume of 1-1.5 mL, via stomach intubation.⁹⁰⁻⁹²

PHARMACOKINETICS OF NAB-PACLITAXEL-LOADED PLGA NANOPARTICLE

According to Stage et al., the most commonly used dosage regimen for paclitaxel is 175 mg/m² with a 3-h infusion. This popular dosage regimen is correlated with a median CL of 12 L/h/m² and a maximum concentration (C_{max}) of 5 mmol/L.⁹³ Additionally, the half-life of paclitaxel is estimated to be 6-13 h after intravenous administration.⁹⁴

Paclitaxel is commonly administered intravenously; therefore, it enters the blood circulation and bypasses gastrointestinal absorption.⁹⁵ Following this, paclitaxel is distributed extensively throughout the body, and its concentration decreases through 2 phases.⁹⁵ The distribution throughout the peripheral body parts is indicated by the rapid decline of paclitaxel concentration, whereas the slow decline in the second phase indicates paclitaxel elimination.³ Paclitaxel is metabolized in the liver to 6 α -hydroxy paclitaxel by cytochrome 2C8, and to two minor metabolites, 3-p-hydroxy paclitaxel as well as 6 α , 3'-p-dihydroxy paclitaxel, by cytochrome 3A4.^{94,95} Paclitaxel is mostly eliminated through biliary excretion and metabolism, whereas renal CL plays a minor role.⁹⁴ Nab-paclitaxel was developed to improve the pharmacokinetics, pharmacodynamics, and safety profile by eliminating the potential toxicity of polyethoxylated castor oil components while the efficacy of paclitaxel was maintained or increased.^{80,95,96} According to Giordano et al., nab-paclitaxel particles have a smaller diameter; therefore, there is an in-

crease in the intracellular distribution of paclitaxel and higher antitumor activity.⁹⁶ Apart from the larger volume of distribution, nab-paclitaxel also has a higher concentration and faster CL rate than conventional paclitaxel.⁹⁶

One of the most effective and potent antitumor agents, nab-paclitaxel poses major challenges, namely extremely poor water solubility (<0.025 mg/mL), possible MDR in some tumor cells, and nonspecific pharmacokinetics in the systemic circulation.⁷⁹ Thus, to overcome these challenges, a nab-paclitaxel delivery system was developed, one of which is PLGA nanoparticles.⁷⁹ PLGA nanoparticles after surface modification, for example, through PEGylation, have the potential to increase blood circulation time and improve drug pharmacokinetics.²² This was supported by an *in vitro* study by Wei et al., who found that curcumin-loaded PLGA synthesized by PEGylation had improved pharmacokinetic properties and increased drug bioavailability by up to 55.4 times.⁷ Rezvantalab et al. concluded that PLGA nanoparticle implementation with the antitumor docetaxel-loaded PLGA accumulated lesser in the liver, spleen, and lungs compared to free docetaxel.^{22,97} Therefore, PLGA nanoparticles demonstrate high potential in improving the pharmacokinetics of nab-paclitaxel as a treatment modality for pancreatic cancer.

PHARMACODYNAMICS OF NAB-PACLITAXEL-LOADED PLGA

Compared with paclitaxel molecules, nab-paclitaxel has a smaller molecular diameter structure, allowing intracellular transport of paclitaxel and better antitumor activity. Albumin binding facilitates the transport of paclitaxel across endothelial cells via the albumin receptor pathway, the gp60 receptor. This increases the number of paclitaxel that perform endothelial transcytosis by 4.2 times in the presence of albumin nanoparticles. The albumin nano bind also interacts with a glycoprotein receptor, secreted protein acidic and cysteine rich/osteonectin that is over-expressed in certain tumor cases, such as pancreatic ductal adenocarcinoma (PDAC), resulting in a much more selective action of nab-paclitaxel on tumor cells.⁹⁶ PLGA is advantageous in the drug delivery aspect of nab-paclitaxel, binding to the gastrointesti-

nal mucosa to extend the duration of drug absorption. The hydrolysis of PLGA results in the formation of lactic acid and glycolic acid, which are endogenous materials that can be utilized in the Krebs cycle.²² Owing to the biocompatible nature of PLGA, it can be modified into various shapes and sizes of desired particles to be compatible with various organic solvents to deliver nab-paclitaxel to specific tumor cells, in this case, PDAC cells.^{22,98}

CLINICAL EFFECTS OF NAB-PACLITAXEL-LOADED PLGA

Nab-paclitaxel-loaded PLGA nanoparticles in the body are determined through the pharmacodynamic mechanisms, as a therapeutic method of therapy to manage pancreatic cancer, to inhibit the mitosis of cancerous cells, which is a therapeutic effect.^{80,99} Paclitaxel exhibited a greater PFS with a result of 12.9 vs. 7.5 months ($p=0.0065$) and a superior ORR in a randomized multicentre study comparing the two first-line treatments in metastatic breast cancer, paclitaxel and docetaxel.^{90,91} In contrast, an early Phase I clinical trial found that nab-paclitaxel has the maximum tolerated dosage of 300 mg/m², which is 70% greater than traditional paclitaxel (175 mg/m²) and hence does not produce harmful side effects.¹⁰⁰

Hersh et al. found that in Phase III clinical trial, the ratio of overall survival (OS) and ORR between dacarbazine and nab-paclitaxel was higher for nab-paclitaxel by 15% and 11%, respectively.¹⁰¹ A Phase II study of 43 patients treated with 260 mg/m² every 3 weeks demonstrated a benefit in 49% of cases and a 16% ORR, with PFS and OS of 6 and 11 months, respectively.¹⁰² Patients in the nab-paclitaxel group had a lower rate of hypersensitivity than those in the conventional group, according to Zong et al.^{80,103} Gradishar et al. found that when nab-paclitaxel was compared to polyethylated castor oil-based paclitaxel, it had considerably greater response rates (33% vs. 19%; $p=0.001$).¹⁰⁴ According to studies done by Xu et al., delivering PLGA nanoparticles with a low molecular weight increases the rate of breakdown and drug release.¹⁰⁵ Furthermore, PLGA nanoparticles loaded with nab-paclitaxel demonstrated a 3.7-fold longer drug elimination half-life than the over-the-counter medications.^{22,97}

In a Phase II trial, the desmoplastic stroma was discovered to be depleted in mice with human pancreatic cancer xenografts treated with nab-paclitaxel alone or in combination with gemcitabine.¹⁰⁶ In continuation to the previous study, it was discovered that the nab-paclitaxel-gemcitabine group exhibited 35% and 9% pancreatic cancer survival rates at 1 and 2 years, respectively, while the gemcitabine group had a pancreatic cancer survival rate of 22% and 4%, respectively. The nab-paclitaxel-gemcitabine group also showed a longer median PFS of 5.5 months compared to 3.7 months in the gemcitabine group.¹⁰⁶ An advanced nanoparticle system has been designed by Massey et al. to target pancreatic cancer by developing a multi-layered formulation of paclitaxel-loaded PLGA nanoparticles coated with poly-L-lysine and stabilized with Pluronic F127 (Bio-Engineering Co. Ltd, Xi'an, China). This formulation demonstrated desirable characteristics such as optimal size (~160 nm) and negative zeta potential (-6.02 mV), effective internalization mediated by lipid rafts, significant inhibition of growth and metastasis in vitro, and both chemo-naïve and chemo-exposed orthotopic xenograft mouse models of pancreatic cancer.²¹ Similarly, Shetty et al. have reported the development of a distinctive paclitaxel-loaded PLGA nanoparticle formulation that is capable of targeting lipid metabolism and augmenting the anticancer activity of chemotherapy drugs in pancreatic cancer cells. Their study found that the paclitaxel-loaded PLGA could effectively inhibit excessive lipid formation and modify membrane stability by reducing the expression of fatty acid synthase, acetyl-CoA carboxylase, lipid, and Cox-2 proteins, as confirmed through Fourier transform infrared and zeta potential measurements.¹⁰⁷ This suggests that the molecular mechanism enhances the efficacy, as demonstrated by its superior inhibitory effects in tumorigenic and metastasis assays in pancreatic cancer cells.

CONCLUSION

Nab-paclitaxel-loaded PLGA nanoparticles are a possible alternative in the treatment of cancer, which currently still has some drawbacks. Nab-paclitaxel-loaded PLGA nanoparticles have a higher endothelial transcytosis ability; therefore, the concentration

of paclitaxel particles to target cells is higher and works more selectively. Nab-paclitaxel-loaded PLGA nanoparticles are constructed using the o/w emulsification solvent evaporation technique and administered intravenously. Nab-paclitaxel-loaded PLGA is extensively distributed and metabolized in cytochrome 2C8, where it is eliminated via biliary excretion. The drug target becomes more selective and undergoes higher endothelial transcytosis in the presence of albumin and PLGA binding. Based on a comparison with other treatment methods using various studies, the nab-paclitaxel-loaded PLGA nanoparticles showed pharmacokinetic and pharmacodynamic advantages.

RECOMMENDATION

Future studies are needed to develop this method, including drug dosage, therapy duration, the presence or absence of mutation effects, possible polypharmacy effects, and the costs involved.

Acknowledgements

The authors would like to express gratitude for colleagues and family for their generous support throughout the literature study.

Source of Finance

During this study, no financial or spiritual support was received neither from any pharmaceutical company that has a direct connection with the research subject, nor from a company that provides or produces medical instruments and materials which may negatively affect the evaluation process of this study.

Conflict of Interest

No conflicts of interest between the authors and / or family members of the scientific and medical committee members or members of the potential conflicts of interest, counseling, expertise, working conditions, share holding and similar situations in any firm.

Authorship Contributions

Idea/Concept: Bryan Gervais de Liyis, Viona Mareska; **Design:** Celine Aurella Ahmad; **Control/Supervision:** Agung Wiewiek Indrayani; **Data Collection and/or Processing:** Celine Aurella Ahmad, Alexandria Cahyaputri Windiarto; **Analysis and/or Interpretation:** Bryan Gervais de Liyis, Viona Mareska; **Literature Review:** Ali Members; **Writing the Article:** Ali Members; **Critical Review:** Agung Wiewiek Indrayani; **References and Fundings:** Alexandria Cahyaputri Windiarto; **Materials:** Bryan Gervais de Liyis.

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. *CA Cancer J Clin.* 2019;69(1):7-34. [[Crossref](#)] [[PubMed](#)]
- McGuigan A, Kelly P, Turkington RC, Jones C, Coleman HG, McCain RS. Pancreatic cancer: A review of clinical diagnosis, epidemiology, treatment and outcomes. *World J Gastroenterol.* 2018;24(43):4846-4861. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Saad AM, Turk T, Al-Husseini MJ, Abdel-Rahman O. Trends in pancreatic adenocarcinoma incidence and mortality in the United States in the last four decades; a SEER-based study. *BMC Cancer.* 2018;18(1):688. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Fazeny FAO. Ikterus obstruktif pada penderita tumor pankreas the obstructive jaundice in patient with pancreatic tumors. *Juni.* 2020;11(1):197-204. [[Crossref](#)]
- Ducreux M, Seufferlein T, Van Laethem JL, et al. Systemic treatment of pancreatic cancer revisited. *Semin Oncol.* 2019;46(1):28-38. [[Crossref](#)] [[PubMed](#)]
- Waters AM, Der CJ. KRAS: The critical driver and therapeutic target for pancreatic cancer. *Cold Spring Harb Perspect Med.* 2018;8(9):a031435. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Buscail L, Bourmet B, Cordelier P. Role of oncogenic KRAS in the diagnosis, prognosis and treatment of pancreatic cancer. *Nat Rev Gastroenterol Hepatol.* 2020;17(3):153-168. [[Crossref](#)] [[PubMed](#)]
- Saung MT, Zheng L. Current standards of chemotherapy for pancreatic cancer. *Clin Ther.* 2017;39(11):2125-2134. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Berdiss AJ. Inhibiting DNA polymerases as a therapeutic intervention against cancer. *Front Mol Biosci.* Nov 2017;4:78. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Hamada C, Okusaka T, Ikari T, et al. Efficacy and safety of gemcitabine plus S-1 in pancreatic cancer: a pooled analysis of individual patient data. *Br J Cancer.* 2017;116(12):1544-1550. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Wu P, Wang X, Ma Y, et al. (3E,5E)-3,5-Bis(pyridin-3-methylene)-tetrahydrothiopyran-4-one enhances the inhibitory effect of gemcitabine on pancreatic cancer cells. *Bioorg Chem.* Aug 2020;101:104022. [[Crossref](#)] [[PubMed](#)]
- Alam S, Illo C, Ma YT, Punia P. Gemcitabine-Induced Cardiotoxicity in Patients Receiving Adjuvant Chemotherapy for Pancreatic Cancer: A Case Series. *Case Rep Oncol.* 2018;11(1):221-227. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Wang Y, Hu GF, Zhang QQ, et al. Efficacy and safety of gemcitabine plus erlotinib for locally advanced or metastatic pancreatic cancer: a systematic review and meta-analysis. *Drug Des Devel Ther.* Jun 2016;10:1961-1972. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Bendell J, Sharma S, Patel MR, et al. Safety and efficacy of andeciximab (GS-5745) plus gemcitabine and nab-paclitaxel in patients with advanced pancreatic adenocarcinoma: results from a phase I study. *Oncologist.* 2020;25(11):954-962. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Morizane C, Okusaka T, Mizusawa J, et al; members of the Hepatobiliary and Pancreatic Oncology Group of the Japan Clinical Oncology Group (JCOG-HBPOG). Combination gemcitabine plus S-1 versus gemcitabine plus cisplatin for advanced/recurrent biliary tract cancer: the FUGA-BT (JCOG1113) randomized phase III clinical trial. *Ann Oncol.* 2019;30(12):1950-1958. [[PubMed](#)]
- Christensen JG, Olson P, Briere T, Wiel C, Bergo MO. Targeting Krasg12c -mutant cancer with a mutation-specific inhibitor. *J Intern Med.* 2020;288(2):183-191. [[Crossref](#)] [[PubMed](#)]
- Salgia R, Pharaon R, Mambetsariev I, Nam A, Sattler M. The improbable targeted therapy: KRAS as an emerging target in non-small cell lung cancer (NSCLC). *Cell Rep Med.* 2021;2(1):100186. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Tischer J, Gergely F. Anti-mitotic therapies in cancer. *J Cell Biol.* 2019;218(1):10-11. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Alqahtani FY, Aleanizy FS, El Tahir E, Alkahtani HM, AlQuadeib BT. Paclitaxel. *Profiles Drug Subst Excip Relat Methodol.* Apr 2019;44:205-238. [[Crossref](#)] [[PubMed](#)]
- Senapati S, Mahanta AK, Kumar S, Maiti P. Controlled drug delivery vehicles for cancer treatment and their performance. *Signal Transduct Target Ther.* 2018;3(1):7. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Massey AE, Sikander M, Chauhan N, Kumari S, Setua S, Shetty AB, et al. Next-generation paclitaxel-nanoparticle formulation for pancreatic cancer treatment. *Nanomedicine.* Aug 2019;20:102027. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Rezvantlab S, Drude NI, Moraveji MK, et al. PLGA-based nanoparticles in cancer treatment. *Front Pharmacol.* Nov 2018;9:1260. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Ghitman J, Biru EI, Stan R, Iovu H. Review of hybrid PLGA nanoparticles: Future of smart drug delivery and theranostics medicine. *Mater Des.* August 2020;193:108805. [[Crossref](#)]
- Wu ST, Fowler AJ, Garmon CB, et al. Treatment of pancreatic ductal adenocarcinoma with tumor antigen specific-targeted delivery of paclitaxel loaded PLGA nanoparticles. *BMC Cancer.* 2018;18(1):457. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Cullis J, Siolas D, Avanzi A, Barui S, Maitra A, Bar-Sagi D. Macropinocytosis of Nab-paclitaxel Drives Macrophage Activation in Pancreatic Cancer. *Cancer Immunol Res.* 2017;5(3):182-190. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Zhou Q, Melton DA. Pancreas regeneration. *Nature.* 2018;557(7705):351-358. Erratum in: *Nature.* 2018;560(7720):E34. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Wynne K, Devereaux B, Dornhorst A. Diabetes of the exocrine pancreas. *J Gastroenterol Hepatol.* 2019;34(2):346-354. [[Crossref](#)] [[PubMed](#)]
- Bakhti M, Böttcher A, Lickert H. Modelling the endocrine pancreas in health and disease. *Nat Rev Endocrinol.* 2019;15(3):155-171. [[Crossref](#)] [[PubMed](#)]
- Zhang L, Sanagapalli S, Stoita A. Challenges in diagnosis of pancreatic cancer. *World J Gastroenterol.* 2018;24(19):2047-2060. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Hata T, Suenaga M, Marchionni L, Macgregor-Das A, Yu J, Shindo K, et al. Genome-wide somatic copy number alterations and mutations in high-grade pancreatic intraepithelial neoplasia. *Am J Pathol.* 2018;188(7):1723-1733. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Zhu B, Zhu Y, Tian J, et al. A functional variant rs1537373 in 9p21.3 region is associated with pancreatic cancer risk. *Mol Carcinog.* 2019;58(5):760-766. [[Crossref](#)] [[PubMed](#)]
- Lim CS, Im K, Lee DS, et al. The implication of cytogenetic alterations in pancreatic ductal adenocarcinoma and intraductal papillary mucinous neoplasm identified by fluorescence in situ hybridization and their potential diagnostic utility. *Gut Liver.* 2020;14(4):509-520. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Collateral Lethality in Pancreatic Cancer. *Cancer Discov.* 2017;7(4):342-343. [[Crossref](#)] [[PubMed](#)]
- Chen YJ, Ojeaburu JV, Vortmeyer A, Yu S, Jensen RT. Alterations of chromosome 3p in 24 cases of gastrinomas and their correlations with clinicopathological and prognostic features. *J Pancreatol.* 2020;3(1):42-49. [[Crossref](#)]
- Vašíčková K, Horak P, Vaňhara P. TUSC3: functional duality of a cancer gene. *Cell Mol Life Sci.* 2018;75(5):849-857. [[Crossref](#)] [[PubMed](#)]
- Driehuis E, van Hoeck A, Moore K, et al. Pancreatic cancer organoids recapitulate disease and allow personalized drug screening. *Proc Natl Acad Sci U S A.* 2019;116(52):26580-26590. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
- Baeeri M, Rahimifard M, Daghighi SM, et al. Cannabinoids as anti-ROS in aged pancreatic islet cells. *Life Sci.* Sep 202 ;256:117969. [[Crossref](#)] [[PubMed](#)]
- Zińczuk J, Zaręba K, Guzińska-Ustymowicz K, Kędra B, Kemona A, Pryczynicz A. p16, p21, and p53 proteins play an important role in development of pancreatic intraepithelial neoplastic. *Ir J Med Sci.* 2018;187(3):629-637. [[Crossref](#)] [[PubMed](#)]

39. Mello SS, Attardi LD. Neat-en-ing up our understanding of p53 pathways in tumor suppression. *Cell Cycle*. 2018;17(13):1527-1535. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
40. Ahmed S, Schwartz C, Dewan MZ, Xu R. The Promising Role of TGF- β /SMAD4 in Pancreatic Cancer: The future targeted therapy. *J Cancer Treat & Diagnosis*. 2019;3(2):1-7. [[Crossref](#)]
41. Douville C, Springer S, Kinde I, et al. Detection of aneuploidy in patients with cancer through amplification of long interspersed nucleotide elements (LINEs). *Proc Natl Acad Sci U S A*. 2018;115(8):1871-1876. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
42. Kim DU. Does the cytogenetic analysis using fluorescence in situ hybridization improve the preoperative diagnostic accuracy of pancreatic ductal adenocarcinoma? *Gut Liver*. 2020;14(4):397-398. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
43. Zafra MP, Parsons MJ, Kim J, et al. An in vivo kras allelic series reveals distinct phenotypes of common oncogenic variants. *Cancer Discov*. 2020;10(11):1654-1671. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
44. Pipinikas CP, Berner AM, Sposito T, Thirlwell C. The evolving (epi)genetic landscape of pancreatic neuroendocrine tumours. *Endocr Relat Cancer*. 2019;26(9):R519-R544. [[Crossref](#)] [[PubMed](#)]
45. Sharif S, Ramanathan RK, Potter D, Cieply K, Krasinskas AM. HER2 gene amplification and chromosome 17 copy number do not predict survival of patients with resected pancreatic adenocarcinoma. *Dig Dis Sci*. 2008;53(11):3026-3032. [[Crossref](#)] [[PubMed](#)]
46. Han SH, Ryu KH, Kwon AY. The prognostic impact of HER2 genetic and protein expression in pancreatic carcinoma-HER2 protein and gene in pancreatic cancer. *Diagnostics (Basel)*. 2021;11(4):653. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
47. Huang Y, Wei J, Fang Y, et al. Prognostic value of AIB1 and EIF5A2 in intravesical recurrence after surgery for upper tract urothelial carcinoma. *Cancer Manag Res*. Dec 2018;10:6997-7011. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
48. Li L, Bao J, Wang H, et al. Upregulation of amplified in breast cancer 1 contributes to pancreatic ductal adenocarcinoma progression and vulnerability to blockage of hedgehog activation. *Theranostics*. 2021;11(4):1672-1689. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
49. Rice A, Del Rio Hernandez A. The mutational landscape of pancreatic and liver cancers, as represented by circulating tumor DNA. *Front Oncol*. Sep 2019;9:952. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
50. Gu Y, Ji Y, Jiang H, Qiu G. Clinical effect of driver mutations of KRAS, CDKN2A/P16, TP53, and SMAD4 in pancreatic cancer: a meta-analysis. *Genet Test Mol Biomarkers*. 2020;24(12):777-788. [[Crossref](#)] [[PubMed](#)]
51. Murugan AK, Grieco M, Tsuchida N. RAS mutations in human cancers: Roles in precision medicine. *Semin Cancer Biol*. Dec 2019;59:23-35. [[Crossref](#)] [[PubMed](#)]
52. Pązik M, Michalska K, Żebrowska-Nawrocka M, Zawadzka I, Łochowski M, Balcerczak E. Clinical significance of HRAS and KRAS genes expression in patients with non-small-cell lung cancer - preliminary findings. *BMC Cancer*. 2021;21(1):130. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
53. Fan Z, Fan K, Yang C, et al. Critical role of KRAS mutation in pancreatic ductal adenocarcinoma. *Transl Cancer Res*. 2018;7(6):1728-1736. [[Crossref](#)]
54. Terrell EM, Morrison DK. Ras-mediated activation of the Raf family kinases. *Cold Spring Harb Perspect Med*. 2019;9(1):a033746. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
55. Pantsar T. The current understanding of KRAS protein structure and dynamics. *Comput Struct Biotechnol J*. Dec 2019;18:189-198. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
56. Simanshu DK, Nissley DV, McCormick F. RAS proteins and their regulators in human disease. *Cell*. 2017;170(1):17-33. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
57. Lechuga CG, Simón-Carrasco L, Jacob HK, Drost M. Genetic validation of cell proliferation via Ras-independent activation of the Raf/Mek/Erk pathway. *Methods Mol Biol*. 2017;1487:269-276. [[Crossref](#)] [[PubMed](#)]
58. Guo YJ, Pan WW, Liu SB, Shen ZF, Xu Y, Hu LL. ERK/MAPK signalling pathway and tumorigenesis. *Exp Ther Med*. 2020;19(3):1997-2007. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
59. Qi C, Wang X, Shen Z, Chen S, Yu H, Williams N, et al. Anti-mitotic chemotherapeutics promote apoptosis through TL1A-activated death receptor 3 in cancer cells. *Cell Res*. 2018;28(5):544-555. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
60. Forth S, Kapoor TM. The mechanics of microtubule networks in cell division. *J Cell Biol*. 2017;216(6):1525-1531. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
61. Bernabeu E, Cagel M, Lagomarsino E, Moreton M, Chiappetta DA. Paclitaxel: What has been done and the challenges remain ahead. *Int J Pharm*. 2017;526(1-2):474-495. [[Crossref](#)] [[PubMed](#)]
62. Zhu L, Chen L. Progress in research on paclitaxel and tumor immunotherapy. *Cell Mol Biol Lett*. Jun 2019;24:40. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
63. Wei Y, Pu X, Zhao L. Preclinical studies for the combination of paclitaxel and curcumin in cancer therapy (Review). *Oncol Rep*. 2017;37(6):3159-3166. [[Crossref](#)] [[PubMed](#)]
64. Miller EM, Samec TM, Alexander-Bryant AA. Nanoparticle delivery systems to combat drug resistance in ovarian cancer. *Nanomedicine*. Jan 2021;31:102309. [[Crossref](#)] [[PubMed](#)]
65. Yao Y, Zhou Y, Liu L, et al. Nanoparticle-based drug delivery in cancer therapy and its role in overcoming drug resistance. *Front Mol Biosci*. Aug 2020;7:193. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
66. Sharifi-Rad J, Quispe C, Patra JK, et al. Paclitaxel: application in modern oncology and nanomedicine-based cancer therapy. *Oxid Med Cell Longev*. Oct 2021;2021:3687700. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
67. Karimi M, Bahrami S, Ravari SB, et al. Albumin nanostructures as advanced drug delivery systems. *Expert Opin Drug Deliv*. 2016;13(11):1609-1623. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
68. Cortes J, Saura C. Nanoparticle albumin-bound (nabTM)-paclitaxel: improving efficacy and tolerability by targeted drug delivery in metastatic breast cancer. *Eur J Cancer Suppl*. 2010;8(1):1-10. [[Crossref](#)]
69. Dent S, Fraser J, Graham N, Campbell M, Hopkins S, Dranitsaris G. Clinical outcomes of women with metastatic breast cancer treated with nab-paclitaxel: experience from a single academic cancer centre. *Curr Oncol*. 2013;20(1):24-29. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
70. Liu M, Liu S, Yang L, Wang S. Comparison between nab-paclitaxel and solvent-based taxanes as neoadjuvant therapy in breast cancer: a systematic review and meta-analysis. *BMC Cancer*. 2021;21(1):118. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
71. Mahtani RL, Parisi M, Glück S, et al. Comparative effectiveness of early-line nab-paclitaxel vs. paclitaxel in patients with metastatic breast cancer: a US community-based real-world analysis. *Cancer Manag Res*. Feb 2018;10:249-256. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
72. Alvi M, Yaqoob A, Rehman K, Shoaib SM, Akash MSH. PLGA-based nanoparticles for the treatment of cancer: current strategies and perspectives. *AAPS Open*. 2022;8(1):12. [[Crossref](#)]
73. Allyn MM, Luo RH, Hellwarth EB, Swindle-Reilly KE. Considerations for polymers used in ocular drug delivery. *Front Med (Lausanne)*. Jan 2022;8:787644. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
74. Duncan R. Polymer conjugates as anticancer nanomedicines. *Nat Rev Cancer*. 2006;6(9):688-701. [[Crossref](#)] [[PubMed](#)]
75. Sharma N, Kumari RM, Gupta N, Syed A, Bahkali AH, Nimesh S. Poly-(lactic-co-glycolic) acid nanoparticles for synergistic delivery of epirubicin and paclitaxel to human lung cancer cells. *Molecules*. 2020;25(18):4243. [[Crossref](#)] [[PubMed](#)] [[PMC](#)]
76. Crosasso P, Ceruti M, Brusa P, Arpicco S, Dosio F, Cattel L. Preparation, characterization and properties of sterically stabilized paclitaxel-containing liposomes. *J Control Release*. 2000;63(1-2):19-30. [[Crossref](#)] [[PubMed](#)]
77. Yoshizawa Y, Kono Y, Ogawara K, Kimura T, Higaki K. PEG liposomalization of paclitaxel improved its in vivo disposition and anti-tumor efficacy. *Int J Pharm*. 2011;412(1-2):132-141. [[Crossref](#)] [[PubMed](#)]

78. Ding D, Zhu Q. Recent advances of PLGA micro/nanoparticles for the delivery of biomacromolecular therapeutics. *Mater Sci Eng C Mater Biol Appl*. Nov 2018;92:1041-1060. [Crossref] [PubMed]
79. Zhang Z, Wang X, Li B, et al. Paclitaxel-loaded PLGA microspheres with a novel morphology to facilitate drug delivery and antitumor efficiency. *RSC Adv*. 2018;8(6):3274-3285. [Crossref] [PubMed] [PMC]
80. Abu Samaan TM, Samec M, Liskova A, Kubatka P, Büsselberg D. Paclitaxel's mechanistic and clinical effects on breast cancer. *Biomolecules*. 2019;9(12):789. [Crossref] [PubMed] [PMC]
81. Gorain B, Choudhury H, Pandey M, Kesharwani P. Paclitaxel loaded vitamin E-TPGS nanoparticles for cancer therapy. *Mater Sci Eng C Mater Biol Appl*. Oct 2018;91:868-880. [Crossref] [PubMed]
82. Alam T, Khan S, Gaba B, Haider MF, Baboota S, Ali J. Nanocarriers as treatment modalities for hypertension. *Drug Deliv*. 2017;24(1):358-369. [Crossref] [PubMed] [PMC]
83. Yetisgin AA, Cetinel S, Zuvun M, Kosar A, Kutlu O. Therapeutic nanoparticles and their targeted delivery applications. *Molecules*. 2020;25(9):2193. [Crossref] [PubMed] [PMC]
84. Khameneh ES, Amini MM, Kakaei S, Khanchi A. Preparation of dual-modality yttrium-90 radiolabeled nanoparticles for therapeutic investigation. *Radiochim Acta*. 2018;106(11):897-907. [Crossref]
85. Zhang M, Merlin D. Nanoparticle-based oral drug delivery systems targeting the colon for treatment of ulcerative colitis. *Inflamm Bowel Dis*. 2018;24(7):1401-1415. [Crossref] [PubMed] [PMC]
86. Liyanage PY, Hettiarachchi SD, Zhou Y, et al Nanoparticle-mediated targeted drug delivery for breast cancer treatment. *Biochim Biophys Acta Rev Cancer*. 2019;1871(2):419-433. [Crossref] [PubMed] [PMC]
87. Mitchell MJ, Billingsley MM, Haley RM, Wechsler ME, Peppas NA, Langer R. Engineering precision nanoparticles for drug delivery. *Nat Rev Drug Discov*. 2021;20(2):101-124. [Crossref] [PubMed] [PMC]
88. Jain KK. An Overview of drug delivery systems. *Methods Mol Biol*. 2020;2059:1-54. [Crossref] [PubMed]
89. Smith SA, Selby LI, Johnston APR, Such GK. The Endosomal Escape of Nanoparticles: Toward More Efficient Cellular Delivery. *Bioconjug Chem*. 2019;30(2):263-272. [Crossref] [PubMed]
90. Zhang E, Xing R, Liu S, Qin Y, Li K, Li P. Advances in chitosan-based nanoparticles for oncotherapy. *Carbohydr Polym*. Oct 2019;222:115004. [Crossref] [PubMed]
91. Kopeckova K, Eckschlager T, Sirc J, Hobzova R, Plch J, Hrabeta J, et al. Nanodrugs used in cancer therapy. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*. 2019;163(2):122-131. [Crossref] [PubMed]
92. Matsui H, Hazama S, Shindo Y, Nagano H. Combination treatment of advanced pancreatic cancer using novel vaccine and traditional therapies. *Expert Rev Anticancer Ther*. 2018;18(12):1205-1217. [Crossref] [PubMed]
93. Borgå O, Lilienberg E, Bjermo H, Hansson F, Heldring N, Dediu R. Pharmacokinetics of total and unbound paclitaxel after administration of paclitaxel micellar or nab-paclitaxel: an open, randomized, cross-over, explorative study in breast cancer patients. *Adv Ther*. 2019;36(10):2825-2837. [Crossref] [PubMed] [PMC]
94. Aronson JK. Meyler's Side Effects of Drugs. Arason JK. The International Encyclopedia of Adverse Drug Reactions and Interactions. 16th ed. Amsterdam: Elsevier Science; 2016. p.445-52.
95. Li Q, Zhang H, Zhu X, et al. Tolerance, variability and pharmacokinetics of albumin-bound paclitaxel in Chinese breast cancer patients. *Front Pharmacol*. Nov 2018;9:1372. [Crossref] [PubMed] [PMC]
96. Giordano G, Pancione M, Olivieri N, et al. Nano albumin bound-paclitaxel in pancreatic cancer: Current evidences and future directions. *World J Gastroenterol*. 2017;23(32):5875-5886. [Crossref] [PubMed] [PMC]
97. Rafiei P, Haddadi A. Docetaxel-loaded PLGA and PLGA-PEG nanoparticles for intravenous application: pharmacokinetics and biodistribution profile. *Int J Nanomedicine*. Jan 2017;12:935-947. [Crossref] [PubMed] [PMC]
98. Jiang G, Jia H, Qiu J, et al. PLGA Nanoparticle platform for trans-ocular barrier to enhance drug delivery: a comparative study based on the application of oligosaccharides in the outer membrane of carriers. *Int J Nanomedicine*. Nov 2020;15:9373-9387. [Crossref] [PubMed] [PMC]
99. Talmadge E King. Paclitaxel (Taxol) | Cancer information | Cancer Research UK. 2019. Accessed on 16 April 2022. [Link]
100. Stage TB, Bergmann TK, Kroetz DL. Clinical pharmacokinetics of paclitaxel monotherapy: an updated literature review. *Clin Pharmacokinet*. 2018;57(1):7-19. [Crossref] [PubMed] [PMC]
101. Specenier P. Efficacy of nab-paclitaxel in treating metastatic melanoma. *Expert Opin Pharmacother*. 2019;20(5):495-500. [Crossref] [PubMed]
102. Adrianzen Herrera D, Ashai N, Perez-Soler R, Cheng H. Nanoparticle albumin bound-paclitaxel for treatment of advanced non-small cell lung cancer: an evaluation of the clinical evidence. *Expert Opin Pharmacother*. 2019;20(1):95-102. [Crossref] [PubMed]
103. Zong Y, Wu J, Shen K. Nanoparticle albumin-bound paclitaxel as neoadjuvant chemotherapy of breast cancer: a systematic review and meta-analysis. *Oncotarget*. 2017;8(10):17360-17372. [Crossref] [PubMed] [PMC]
104. Gradisher WJ, Tjulandin S, Davidson N, et al. Phase III trial of nanoparticle albumin-bound paclitaxel compared with polyethylated castor oil-based paclitaxel in women with breast cancer. *J Clin Oncol*. 2005;23(31):7794-7803. [Crossref] [PubMed]
105. Xu Y, Kim CS, Saylor DM, Koo D. Polymer degradation and drug delivery in PLGA-based drug-polymer applications: A review of experiments and theories. *J Biomed Mater Res B Appl Biomater*. 2017;105(6):1692-1716. [Crossref] [PubMed]
106. Von Hoff DD, Ramanathan RK, Borad MJ, et al. Gemcitabine plus nab-paclitaxel is an active regimen in patients with advanced pancreatic cancer: a phase I/II trial. *J Clin Oncol*. 2011;29(34):4548-4554. [Crossref] [PubMed] [PMC]
107. Shetty A, Nagesh PKB, Setua S, et al. Novel Paclitaxel Nanoformulation impairs de novo lipid synthesis in pancreatic cancer cells and enhances gemcitabine efficacy. *ACS Omega*. 2020;5(15):8982-8991. [Crossref] [PubMed] [PMC]